

Modulation of Nitrogen Dioxide Toxicity by Lithium¹

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MESSIHA, F S AND J McGRATH *Modulation of nitrogen dioxide toxicity by lithium* PHARMACOL BIOCHEM BEHAV 21: Suppl 1, 93-97, 1984 —The effect of short-term intake of LiCl in drinking fluid on NO₂ toxicity was studied in mice as a function of mortality and of specific activities of mouse liver alcohol dehydrogenase (L-ADH) and aldehyde dehydrogenases (L-ALDH) Pretreatment with LiCl for 10 days decreased mortality in mice exposed to 60 to 70 PPM NO₂ for 6 hr compared to controls Pretreatment with LiCl for 10 days under continued exposure to 5 PPM NO₂ resulted in a decrease in liver weight compared to control Lithium treated mice exposed to NO₂ showed less gain in body weight than the controls treated with LiCl and exposed to air The latter group showed an induction of mitochondrial but not cytoplasmic L-ALDH and the NO₂ exposure did not alter endogenous L-ALDH from corresponding controls This induction of mitochondrial ALDH was associated with an increase in both V_{max} and the apparent K_m Exposure to NO₂ for 10 consecutive days resulted in inhibition of cytoplasmic L-ALDH The data suggest that Li⁺ antagonized NO₂ toxicity A possible mechanism for reduction of NO₂ toxicity by LiCl may be due to Li⁺ action on stabilizing cell membranes and/or modifying intercellular pulmonary response to NO₂ injury

Aldehyde dehydrogenase Lithium Mortality Nitrogen dioxide

INCREASED human exposure to nitrogen oxides (NO_x) from fossil fuel combustion, as well as tobacco smoking, has caused concern that NO₂ may interact with other materials inhaled or ingested by man. Acute exposure to NO₂ is associated with pulmonary edema which is ascribed to increased capillary permeability [15, 17, 35, 39]. Lithium (Li) has been shown to stabilize cellular membranes in various organ systems [6, 12, 37]. This suggests the possibility that Li⁺ may protect against NO₂-induced cellular permeability and reduce morbidity and mortality. This study was conducted to evaluate the effect of short-term pretreatment with LiCl on acute NO₂ toxicity in mice, a species known to be sensitive to NO₂. The effect of LiCl on endogenous hepatic ethanol and acetaldehyde metabolizing enzymes was also studied as a function of NO₂ exposure. This was prompted by the wide misuse of alcohol in general and by changes in activities of these enzymes evoked by Li under normal environmental conditions [26]

METHOD

The subjects were adult albino mice of both sexes obtained from Sprague-Dawley Co., Madison, WI. In the first set of experiments, the effect of pretreatment with LiCl on NO₂ induced mortality was studied. Adult male mice received distilled water or 0.2% LiCl solution as the sole drinking fluid ad lib for 10 consecutive days. They were then challenged for 6 hr with either 60 or 70 PPM NO₂. The LD₅₀ determined in an earlier study was in this range. The number of animals dying in each group immediately and 24 hr after NO₂ exposure was recorded and expressed as per-

cent mortality. The data were analyzed by Chi square analysis.

In the second set of experiments the effect of LiCl on hepatic alcohol dehydrogenase (L-ADH) and aldehyde dehydrogenase (L-ALDH) was studied as a function of exposure to NO₂. Forty adult female mice were divided into two groups of 20 and given either water or 0.2% LiCl as the only drinking fluid. Each group was divided into two subgroups of ten mice each and housed in environmental chambers and exposed to either filtered air or 5 PPM NO₂. The LiCl treatment was initiated concomitant with the NO₂ exposure and lasted for ten consecutive days. Animals were sacrificed by decapitation 20 min post termination of the ten day NO₂ exposure period. The liver was removed quickly and weighed prior to homogenization in 0.1 M KCl buffer pH 6.8. The homogenate was fractionated into subcellular mitochondrial (MT) and cytoplasmic (CT) fractions [27] for the enzymatic assays of L-ALDH [3] and L-ADH [4], respectively. Body weight was determined before and after termination of the experiment. The enzymatic activity was expressed as specific activity, nMol/min/mg protein, measured at 30°C. The kinetics was performed by the Lineweaver and Burk method [23]. Statistical significance of the results was assessed by Student's *t*-test.

The inhalation chambers were constructed of glass and stainless steel and had a volume of 3.24 M³. Filtered air was drawn into each chamber by an electric blower. The chamber system has been detailed elsewhere [24]. The chamber's NO₂ concentration, temperature, pressure and relative humidity were monitored continuously. The airflow was 800 L or 16.0 air changes/hr, at 21±2°C and relative

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TABLE 1
EFFECT OF PRETREATMENT WITH LiCl ON NO₂-PRODUCED MORTALITY IN THE ADULT MALE MOUSE

NO ₂ Concentration (PPM)	Percent Mortality			
	Immediately post NO ₂ inhalation		24 hr post NO ₂ inhalation	
	Controls	Li-treated	Controls	Li-treated
60	67	33	67	50
70	67	17	100	33*

The LiCl was dissolved in distilled water (0.2%) and provided ad lib for 10 consecutive days, (controls received distilled water). Animals were exposed to 60–70 PPM NO₂ for 6 hr. Percent mortality was determined immediately, or 24 hr after termination of the NO₂ exposure. Each treatment group consisted of 6 mice.
* $p < 0.05$

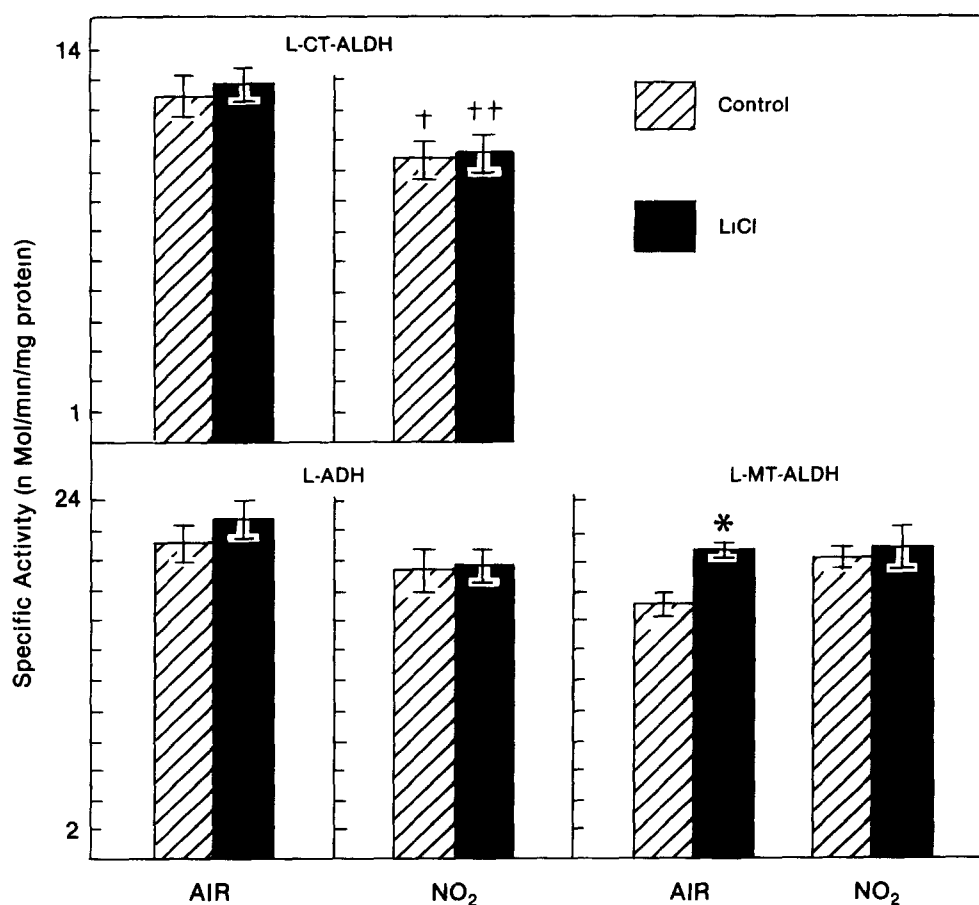


FIG 1 The effect of short-term intake of LiCl on endogenous mouse liver alcohol dehydrogenase (L-ADH) and aldehyde dehydrogenase (L-ALDH) as a function of NO₂ inhalation. Animals breathed 5 PPM NO₂ or filtered air for 10 consecutive days. They received 0.2% LiCl or distilled water ad lib. The L-ALDH was assayed both in cytoplasmic (CT) and in mitochondrial (MT) preparations. Values are means of specific activity \pm SEM of the mean for 10 independent determinations. †† $p < 0.01$, Different from Li-treated mice inhaling air. † $p < 0.05$, Different from drug-free mice inhaling air. * $p < 0.05$, Different from drug-free controls inhaling air.

TABLE 2
EFFECT OF SHORT-TERM EXPOSURE TO NO₂ ON MOUSE BODY AND LIVER WEIGHTS AS A
FUNCTION OF TREATMENT WITH LiCl

NO ₂ Concentration	Drinking Fluid	Body Weight (g)		Liver Weight (g)
		Initial	Terminal	Terminal
0	H ₂ O	28.2 ± 0.9	29.6 ± 0.9	1.348 ± 0.018
0	LiCl/H ₂ O	29.2 ± 1.2	28.6 ± 0.7	1.298 ± 0.064
5 PPM	H ₂ O	26.6 ± 1.5	27.2 ± 1.5	1.345 ± 0.067
5 PPM	LiCl/H ₂ O	26.8 ± 0.7	25.9 ± 0.2†	1.141 ± 0.046*

Adult female mice were exposed to 5 PPM NO₂ for 24 hr, for 10 consecutive days. Controls were exposed to filtered air. Animals received distilled water or 0.2% (w/v) LiCl solution ad lib. Values are means ± SE of the mean of 5 independent determinations.

*Different from drug-free mice inhaling NO₂, $p < 0.05$

†Different from air-inhaling mice treated with LiCl, $p < 0.05$

humidity of $48 \pm 20\%$. The chamber's NO₂ concentration was produced by mixing NO₂ from heated cylinders (Scientific Gas Products) with air by means of a mass flow controller. The NO₂ concentration was determined daily with a chemiluminescence analyzer (Thermolectron) verified by the spectrophotometric method of Saltzman [33]. The analyzer was calibrated weekly by gas-phase titration.

RESULTS

Table 1 summarizes the effects of 6 hr exposure to NO₂ (60–70 PPM) as a function of pretreatment with LiCl. A 67% mortality was recorded in controls immediately after exposure to 60 or 70 PPM NO₂ compared to 33% and 17% ($p < 0.1$) in the LiCl pretreated mice, respectively. Additional death occurred in the group inhaling 70 PPM during the subsequent 24 hr period. Total mortality at this time was 100% in controls and 33% in the Li-treated mice ($p < 0.025$). The Li-pretreatment exerted a protective action against the 60 PPM NO₂ exposure which was not statistically significant.

The effect of LiCl treatment on mouse body and liver weights during NO₂ inhalation for 10 days is given in Table 2. The Li treatment had little effect on body or liver weight in animals breathing filtered air. However, there was a small but a statistically significant ($p < 0.05$) decrease in both body and liver weights in Li-treated mice breathing NO₂ compared to their controls inhaling air. Treatment with LiCl during NO₂ exposure resulted in a decrease in liver weight of approximately 15% ($p < 0.05$) compared to drug-free mice breathing the same NO₂ concentration.

Figure 1 shows the effect of administration of LiCl for 10 consecutive days on mouse L-ADH and L-ALDH as a function of exposure to 5 PPM NO₂ during drug treatment. The Li-treatment induced mitochondrial but not cytoplasmic L-ALDH by approximately 21% ($p < 0.05$) in mice housed under air. This effect was not apparent in Li-treated mice breathing NO₂. Conversely, L-CT-ALDH was inhibited by 17.7% ($p < 0.05$) in drug-free mice breathing NO₂. This inhibition was present also after the Li-treatment ($p < 0.01$). No changes in specific activity of L-ADH were noted under any of experimental conditions used.

Figure 2 illustrates the reciprocals of the velocity of the reaction of L-MT-ALDH against substrate concentration as a function of LiCl treatment in mice breathing filtered air.

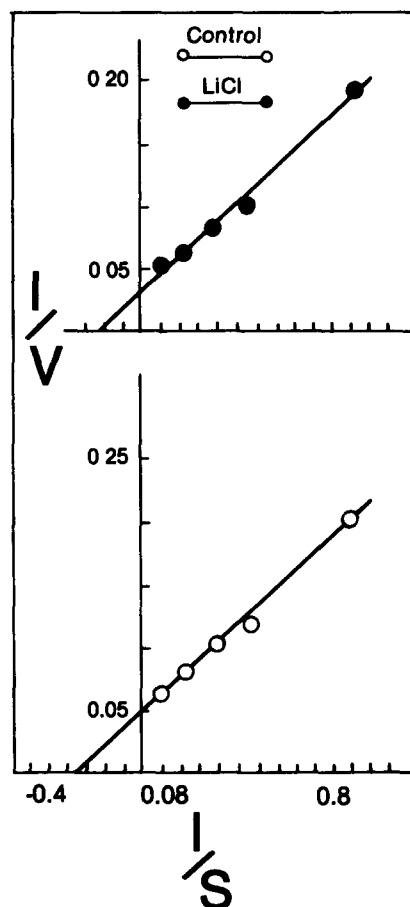


FIG 2 The effect of short-term treatment with LiCl on kinetics of mouse liver mitochondrial aldehyde dehydrogenase. Mice breathed filtered air and received 0.2% LiCl ad lib for 10 days. Reciprocal value of the velocity of the reaction ($1/v$) is plotted against substrate concentration ($1/S$).

The V_{\max} increased from 21.6 to 31.9 units after the Li-treatment. Concomitantly, there was an increase in the apparent K_m from 3.9 in controls to 5.9 units in LiCl treated animals.

DISCUSSION

This study demonstrates that treatment with LiCl for 10 days prior to exposure to high concentrations of NO_2 (60 and 70 PPM, for 6 hr) protected against NO_2 -induced mortality. This Li effect may be of clinical relevance if it occurred in man. For example, NO_2 toxicity is associated with pulmonary dysfunction, respiratory infection and increased susceptibility to lung tumor development [5, 7-9, 11, 13, 14, 30, 36]. Noteworthy, Li-salts possess certain clinical potential in counteracting some of these symptoms. Lithium decreases the incidence of infection by various agents [21, 22, 38] causes mild leukocytosis [32], interferes with certain tumor growth [18] and ameliorates asthma attacks [29,31]. Other immunological actions have been reviewed recently [10]. It is possible that the pulmonary effects evoked by Li^+ may have minimized the lethal effects of high concentration of NO_2 . Moreover, it is likely that the Li-stabilizing effects on cellular membrane [6, 12, 37] may have counteracted the

NO_2 -mediated increase in vascular capillary permeability and edema formation [15, 17, 35, 39].

The mechanism(s) underlying NO_2 toxicity and its modulation by short-term pretreatment with LiCl, remains to be elucidated. Experimental studies on the lethal effect of acute exposure to high concentrations of NO_2 have been focused primarily on pathological changes in the pulmonary system [16, 28, 42]. One biochemical mechanism advanced has been related to NO_2 -induced decrease in pulmonary lipid content [1,20] due to intercellular formation of free radicals [34, 39, 41]. This can be partially prevented by pretreatment with antioxidants [40]. The NO_2 -produced decrease in catalase [41] would tend to support this assumption. Reduced catalase activity decreases detoxification of peroxides and other free radicals. These compounds may accumulate and augment cellular toxicity. Therefore, it appears likely that the NO_2 inhibition of L-C-ALDH noted in this study may reflect derangement in some NAD-dependent dehydrogenases as noted for lactate dehydrogenase [2, 9, 25]. Lithium produced induction of L-MT-ALDH is consistent with previous reports obtained after prolonged ingestion of LiCl by the mouse [26]. In conclusion, Li salts may conceivably have a prophylactic action against NO_2 toxicity.

REFERENCES

- Arner, E. C. and R. A. Rhoades. Long-term nitrogen dioxide exposure. Effects on lung lipids and mechanical properties. *Arch Environ Health* 26: 156-160, 1973.
- Balchum, O. J., R. D. Buckley, R. Sherwin and M. Gardner. Nitrogen dioxide inhalation and lung antibodies. *Arch Environ Health* 10: 274-277, 1965.
- Blair, A. H. and F. H. Bodly. Human liver aldehyde dehydrogenase. Partial purification and properties. *Can J Biochem* 47: 265-272, 1969.
- Blair, A. H. and B. L. Vallee. Some catalytic properties of human liver alcohol dehydrogenase. *Biochemistry* 5: 2026-2034, 1966.
- Cooper, W. C. and I. R. Tabershaw. Biologic effects of nitrogen dioxide in relation to air quality standards. *Arch Environ Health* 12: 522-530, 1966.
- Dunner, D. L., H. L. Meltzer and R. R. Fieve. Metabolism of lithium in erythrocytes. *Mclean Hosp J* 3: 89-97, 1978.
- Ehrlich, R. Effect of air pollutants on respiratory infection. *Arch Environ Health* 6: 638-642, 1963.
- Ehrlich, R. Effect of nitrogen dioxide on resistance to respiratory infection. *Bacteriol Rev* 30: 604-614, 1966.
- Ehrlich, R. and M. C. Henry. Chronic toxicity of nitrogen dioxide. I. Effect on resistance to bacterial pneumonia. *Arch Environ Health* 17: 860-865, 1968.
- Frost, R. E. and F. S. Messiha. Clinical uses of lithium salts. *Brain Res Bull* 11: 219-231, 1983.
- Gardner, M. B. Biological effects of urban air pollution. III. Lung tumors in mice. *Arch Environ Health* 12: 305-313, 1966.
- Glen, A. I. M. The effects of lithium on cell membranes. *Ecerpta Med Int Congr Ser* 478: 768-780, 1979.
- Goldstein, E. Evaluation of the role of nitrogen dioxide in the development of respiratory diseases in man. *Calif Med* 115: 21-27, 1971.
- Goldstein, E., M. C. Eagle and P. D. Hoepflich. Effects of nitrogen dioxide on pulmonary bacterial defense mechanism. *Arch Environ Health* 26: 202-204, 1973.
- Gregory, A. R. Inhalation toxicology and lung edema receptor sites. *Am Ind Hyg Assoc J* 31: 454-459, 1970.
- Henry, M. C., R. Ehrlich and W. H. Blair. Effect of nitrogen dioxide on resistance of squirrel monkeys to *Klebsiella pneumoniae* infection. *Arch Environ Health* 18: 580-587, 1969.
- Hine, C. H., F. H. Meyers and R. W. Wright. Pulmonary changes in animals exposed to nitrogen dioxide effects of acute exposure. *Toxicol Appl Pharmacol* 16: 201-213, 1970.
- Holler, A. C. and R. V. Huch. Colorimetric determination of nitrates and nitric acid esters. Isometric xylenols as reagents. *Anal Chem* 21: 1385-1389, 1969.
- Ito, K., K. Motomiya, R. Yoshida, H. Otsu and T. Nakajima. Effect of nitrogen dioxide inhalation on influenza virus in mice. *Jpn J Hyg* 26: 304-314, 1971.
- Kensler, C. J. and S. P. Battista. Components of cigarette smoke with ciliary depressant-activity. Their selective removal by filters containing activated charcoal granules. *N Engl J Med* 269: 1161-1166, 1963.
- Lapierre, G. and R. B. Stewart. Lithium carbonate and leukocytosis. *Am J Hosp Pharm* 37: 1525-1528, 1980.
- Lee, M. and L. E. Hoppins. Attenuation of chemotherapy-induced neutropenia with lithium carbonate. *Am J Hosp Pharm* 37: 1066-1071, 1980.
- Lineweaver, H. and D. Burk. Determination of enzyme dissociation constants. *J Am Chem Soc* 56: 658-666, 1934.
- McGrath, J. and J. Oyervides. Response of NO_2 -exposed mice to *Klebsiella* challenge. *Adv Mod Environ Toxicol* 5: 475-485, 1983.
- Messiha, F. S., J. McGrath, J. Early, M. J. Hughes and D. F. Rector. Biochemical and morphological aspects of nitrogen dioxide toxicity and the effect of ethanol intake. *J Environ Sci Health* 18: 571-581, 1983.
- Messiha, F. S., R. E. Frost and H. F. Sproat. Behavioral, metabolic and histological aspects of lithium and ethanol interaction. *Drug Chem Toxicol* 6: 397-408, 1983.
- Messiha, F. S. and M. J. Hughes. Liver alcohol and aldehyde dehydrogenase. Inhibition and potentiation by histamine agonists and antagonists. *Clin Exp Pharmacol Physiol* 6: 281-292, 1979.
- Murphy, S. D., C. E. Ulrich, S. H. Frankowitz and C. Xintaras. Altered function in animals inhaling low concentrations of ozone and nitrogen dioxide. *Am Ind Hyg Assoc J* 25: 246-253, 1964.
- Nasr, S. J. and R. W. Atkins. Coincidental improvement in asthma during lithium treatment. *Am J Psychiatry* 134: 1042-1043, 1977.

- 30 Purvis, M R , S Miller and R Ehrlich. Effect of atmospheric pollutants on susceptibility to respiratory infection *J Infect Dis* **109**: 238-242, 1961
- 31 Putmen, P L Possible side effects of lithium *Am J Psychiatry* **135**: 388, 1978
- 32 Risetto, G and G Gassano Variazioni del sangue perifico nella intossicazione spermentale da sali litio *Riv Patal Clin Sper* **7**: 202, 1952
- 33 Saltzman, B E Selected methods for the measurement of air pollutants U S Department of Health Educ & Welfare Public Health Service Publ 999-AP-11, 1965
- 34 Shakman, R A Nutritional influences on the toxicity of environmental pollutants A Review *Arch Environ Health* **28**: 105-113, 1974
- 35 Sherwin, R P and V Richters Lung capillary permeability Nitrogen dioxide exposure and leakage of tritiated serum *Arch Intern Med* **128**: 61-68, 1971
- 36 Shy, C M , J P Creason, M E Pearlman, K E McClain, F B Benson and M M Young The Chattanooga School Children Study Effects of community exposure to nitrogen dioxide Methods description of pollutant exposure and results of ventilatory function testing *J Air Pollut Control Assoc* **20**: 539-545, 1970
- 37 Singer, R. and D Rotenberg Mechanisms of lithium action *N Engl J Med* **72**: 254-260, 1973
- 38 Spina, M P., M Ventura, G. Lavecchia, R Bombara, B Giraldi, F Mensi, A Massari, F. Rossi, G Santi and U Visca Protective effect of lithium carbonate in antitubercular-induced neutropenia and assessment of its long-term effectiveness *Min Med*, **72**: 3323-3328, 1981
- 39 Stokinger, H E and D L Coffin Biological effects of air pollutants In *Air Pollution and Its Effects*, edited by A C Stern New York Academic Press, 1968, pp 445-546.
- 40 Thomas, H V , P K Mueller and R L Lyman Lipoperoxidation of lung lipids in rats exposed to nitrogen dioxide *Science* **159**: 532-534, 1968
- 41 Vanallo, C L , B M Domm, R H Poe, M L Duncombe and J B L Gee NO₂ gas and NO₂ effects on alveolar macrophage phagocytosis and metabolism *Arch Environ Health* **26**: 270-274, 1973
- 42 Von Niedig, G , H Krekeler, R Fuchs, M Wagner and K Koppenhagen Studies of the effects of NO₂ on lung function. Influence on diffusion, perfusion and ventilation in the lungs *Int Arch Arbeitsmed* **31**: 61-72, 1973